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# A Comparative Analysis of Diagnostic Tests for Scrub Typhus

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## Abstract

The Scrub typhus is a prevalent infectious disease caused by Orientia tsutsugamushi, transmitted through mite bites. Prompt and accurate diagnosis is crucial for timely treatment and effective management of the disease. This research abstract provides a comprehensive review and comparative analysis of the diagnostic tests commonly employed for scrub typhus. The primary diagnostic method for scrub typhus is serological testing, which detects antibodies produced in response to the infection. The gold standard test, Indirect Immunofluorescence Assay (IFA), demonstrates high specificity and sensitivity and can confirm the diagnosis by detecting antibody titer rise between acute and convalescent samples. Enzyme-Linked Immunosorbent Assay (ELISA) is an alternative serological test that offers ease of use and cost-effectiveness, although it may have slightly lower sensitivity and specificity. In addition to serological tests, Polymerase Chain Reaction (PCR) has emerged as a valuable diagnostic tool, enabling direct detection of bacterial DNA in various clinical samples. PCR is particularly useful in early-stage infections when antibody levels may be low, although it requires specialized laboratory facilities. The Weil-Felix test, an older diagnostic method, detects antibodies against certain strains of rickettsial bacteria, including Orientia tsutsugamushi. However, due to its limitations in specificity and sensitivity, it is less commonly used in current practice. The comparative analysis highlights the advantages and limitations of each diagnostic test, emphasizing the importance of considering clinical symptoms, patient history, and epidemiological factors in conjunction with test results for accurate diagnosis. False negatives and false positives may occur with any diagnostic method, underscoring the need for consultation with healthcare professionals for appropriate interpretation and subsequent treatment.

**Keywords:** Scrub typhus, Orientia tsutsugamushi, Serological tests, Polymerase Chain Reaction (PCR), Weil-Felix test:



## Introduction

Scrub typhus, a highly prevalent infectious disease, is caused by the bacterium Orientia tsutsugamushi and is primarily transmitted through mite bites [1], [2]. This condition belongs to the group of diseases known as rickettsioses, which are caused by intracellular bacteria. Scrub typhus is endemic to various regions in the Asia-Pacific, including rural areas of Southeast Asia, India, Japan, and parts of Australia [3], [4]. It is considered an important public health issue due to its widespread occurrence and potential complications if left untreated.

The etiological agent of scrub typhus, Orientia tsutsugamushi, is an obligate intracellular bacterium that belongs to the family Rickettsiaceae. This bacterium is unique to scrub typhus and is transmitted to humans through the bite of infected larval mites, commonly known as chiggers or trombiculid mites [5]. These mites acquire the bacteria by feeding on infected vertebrate hosts, including small mammals like rodents [6]. Once infected, the mites can transmit the bacteria to humans during their feeding process.

The clinical presentation of scrub typhus can vary widely, ranging from mild flu-like symptoms to severe and potentially fatal complications. The initial symptoms often include fever, headache, muscle pain, and a characteristic eschar or "tache noire" at the site of the mite bite [7]. This eschar is a localized necrotic skin lesion and is considered a hallmark feature of scrub typhus.

As the disease progresses, patients may develop a variety of systemic manifestations. These can include lymphadenopathy (enlarged lymph nodes), rash, cough, respiratory distress, gastrointestinal symptoms (such as abdominal pain, nausea, and vomiting), and neurological abnormalities. In severe cases, complications like pneumonia, acute respiratory distress syndrome (ARDS), acute kidney injury, myocarditis, and encephalitis can occur, which may increase the mortality rate if not promptly treated [8].

Diagnosing scrub typhus can be challenging, as the symptoms can mimic other febrile illnesses and the availability of diagnostic facilities may be limited in endemic areas. Laboratory tests such as serological assays, polymerase chain reaction (PCR), and immunohistochemistry are used to confirm the diagnosis. Serological tests, such as the indirect immunofluorescence assay (IFA), are commonly employed to detect specific antibodies against Orientia tsutsugamushi in the patient's blood [9].

Prompt and appropriate treatment is crucial in managing scrub typhus to prevent complications and reduce mortality. The antibiotic of choice for scrub typhus is doxycycline, a broad-spectrum antibiotic that is highly effective against the causative agent. Other antibiotics such as azithromycin, clarithromycin, and chloramphenicol are alternatives for patients who cannot tolerate doxycycline. Supportive care, including fluid replacement, management of respiratory distress, and treatment of specific complications, should also be provided as necessary.

Prevention of scrub typhus primarily involves measures aimed at reducing exposure to mite bites. This includes wearing protective clothing, using insect repellents, and employing insecticide-treated bed nets. Vector control measures, such as environmental modification and insecticide spraying, can also be employed to reduce the mite population in endemic areas [10].



This research aims to evaluate and compare the performance of different diagnostic methods in terms of their specificity, sensitivity, ease of use, cost-effectiveness, and other relevant factors. The research specifically focuses on serological tests, such as Indirect Immunofluorescence Assay (IFA) and Enzyme-Linked Immunosorbent Assay (ELISA), which detect antibodies produced in response to the infection. It also explores the utility of Polymerase Chain Reaction (PCR), which allows for the direct detection of bacterial DNA, and the older Weil-Felix test [11].

## **Diagnostic Tests for Scrub Typhus**

Serological tests are crucial diagnostic tools used to identify and monitor infectious diseases, including scrub typhus [12], [13]. These tests detect antibodies produced by the immune system in response to an infection and play a significant role in diagnosing past or present infections and evaluating the immune response to vaccines. Serological testing involves analyzing a blood sample in a laboratory to detect two types of antibodies: immunoglobulin M (IgM) and immunoglobulin G (IgG). IgM antibodies are produced early in the infection, while IgG antibodies persist for a longer duration. Serological tests have extensive applications, aiding in the diagnosis of viral and bacterial infections such as hepatitis, HIV, and Lyme disease. They also contribute to monitoring the spread of infectious diseases, assessing population immunity, and evaluating vaccination effectiveness. For scrub typhus, two commonly used serological tests are the Indirect Immunofluorescence Assay (IFA) and the Enzyme-Linked Immunosorbent Assay (ELISA). These tests provide valuable insights into the presence and concentration of antibodies against Orientia tsutsugamushi, the bacterium responsible for scrub typhus. Another diagnostic method, Polymerase Chain Reaction (PCR), enables the direct detection of bacterial DNA in clinical samples, offering an alternative approach for early diagnosis when antibody levels are low. Although the Weil-Felix test was historically used for rickettsial infections, including scrub typhus, its sensitivity and specificity limitations have led to its decreased use in modern clinical practice in favor of more accurate and specific methods [14], [15].

#### Serological Tests:

Serological tests are the most commonly used diagnostic method for scrub typhus. They detect antibodies produced by the body in response to the infection. Serological tests, also known as antibody tests, are an essential tool in diagnosing and monitoring infectious diseases. These tests detect the presence of antibodies in the blood, which are produced by the immune system in response to an infection. Serological tests play a crucial role in identifying past or present infections and assessing the immune response to vaccines. The process of serological testing involves collecting a blood sample from an individual and analyzing it in a laboratory. The two main types of antibodies detected in serological tests are immunoglobulin M (IgM) and immunoglobulin G (IgG). IgM antibodies are the first to be produced by the immune system in response to an infection, while IgG antibodies are produced later and typically persist for a longer time. By measuring the levels of these antibodies, healthcare professionals can determine if an individual has been exposed to a specific pathogen. Serological tests have various applications in healthcare. They are commonly used for the diagnosis of viral and bacterial infections such as hepatitis, HIV, and Lyme disease. These tests are particularly valuable when other diagnostic methods, such as



PCR tests, are not readily available or when testing for past infections. Serological tests also provide valuable information for public health officials and researchers in monitoring the spread of infectious diseases, assessing population immunity, and evaluating the effectiveness of vaccination campaigns.

The two main types of serological tests used for scrub typhus are Indirect Immunofluorescence Assay (IFA), and Enzyme-Linked Immunosorbent Assay (ELISA) [16], [17].

#### a. Indirect Immunofluorescence Assay (IFA):

Indirect Immunofluorescence Assay (IFA) is a widely used diagnostic test that plays a crucial role in the detection of antibodies against Orientia tsutsugamushi in a patient's blood [16], [18]. It is considered the gold standard for diagnosing scrub typhus, a potentially life-threatening disease caused by the bacterium Orientia tsutsugamushi. The IFA test utilizes specific fluorescently labeled antibodies to detect and visualize the presence of antibodies produced by the patient's immune system in response to the infection.

The IFA test involves several steps to determine the antibody titer against Orientia tsutsugamushi. Firstly, the patient's blood sample is collected and mixed with a known antigen derived from the bacterium. If the patient has been exposed to the bacterium, their immune system produces specific antibodies, which will bind to the antigen [19]. In the next step, a secondary antibody labeled with a fluorescent dye is added. This secondary antibody specifically recognizes and binds to the patient's antibodies that have attached to the antigen [18].

After the addition of the fluorescently labeled secondary antibody, the sample is examined under a fluorescence microscope. If the patient's blood contains antibodies against Orientia tsutsugamushi, they will be visualized as bright fluorescent signals. The intensity of the fluorescence is proportional to the antibody titer, providing information about the immune response and the stage of infection [20].

One advantage of the IFA test is its high sensitivity, allowing for the detection of low levels of antibodies in the patient's blood. This is particularly important during the early stages of infection when antibody levels might be low. Additionally, the IFA test can also provide valuable information about the stage of infection [21]. By comparing the antibody titer in acute and convalescent samples collected weeks apart, a rise in antibody levels confirms the diagnosis of scrub typhus [22].

However, it's important to note that the IFA test requires specialized laboratory equipment, skilled technicians, and proper handling of the samples. The process can be time-consuming, and the interpretation of the results requires expertise. False positives and false negatives can occur due to various factors, such as technical errors or cross-reactivity with antibodies against related organisms.

#### b. Enzyme-Linked Immunosorbent Assay (ELISA):

Enzyme-Linked Immunosorbent Assay (ELISA) is a widely used serological test for the detection of specific antibodies against Orientia tsutsugamushi [18], [23]. It provides a reliable and cost-effective alternative to the Indirect Immunofluorescence Assay (IFA). ELISA involves the use of specific antigens derived from Orientia tsutsugamushi, which are immobilized onto a solid surface, such as a microplate well.



The ELISA test begins by adding the patient's blood sample to the antigen-coated wells. If the patient has been exposed to Orientia tsutsugamushi and has developed antibodies against it, these antibodies will bind to the immobilized antigens. After washing away unbound components, a secondary antibody linked to an enzyme, such as horseradish peroxidase or alkaline phosphatase, is added. This secondary antibody recognizes and binds specifically to the patient's antibodies that are bound to the antigen [24].

In the final step, a substrate specific to the enzyme is added, which, upon interaction with the enzyme, produces a detectable signal, such as a color change. The intensity of the signal is directly proportional to the amount of bound antibodies and can be measured using a spectrophotometer or visually assessed. A higher signal indicates a higher concentration of antibodies against Orientia tsutsugamushi in the patient's blood.

Although ELISA is easier to perform and less expensive than IFA, it may have slightly lower sensitivity and specificity. Sensitivity refers to the ability of the test to correctly identify positive cases, while specificity refers to its ability to correctly identify negative cases [25]. False negatives and false positives can occur due to various factors, including the cross-reactivity of antibodies with other related organisms or technical limitations of the assay. However, ELISA has several advantages, including its scalability, which allows for high-throughput testing, making it suitable for large-scale screening or epidemiological studies [26]. It also has a shorter turnaround time compared to IFA. ELISA can be used to detect antibodies at different stages of infection, including early seroconversion when antibody levels are low.

#### Polymerase Chain Reaction (PCR):

Polymerase Chain Reaction (PCR) tests are powerful molecular techniques that enable the direct detection of bacterial DNA in clinical samples, such as blood, eschar swabs, or tissues [27]. PCR is a valuable tool for diagnosing Orientia tsutsugamushi infections, especially in the early stages of infection when antibody levels may still be low or undetectable.

The PCR test relies on the ability of specific DNA primers to recognize and bind to target sequences within the Orientia tsutsugamushi genome [28]. The clinical sample is processed to extract the bacterial DNA, which serves as the template for the PCR amplification [29]. The extracted DNA is mixed with the PCR reaction mixture containing the primers, DNA polymerase, and nucleotides [30]. The reaction mixture undergoes a series of heating and cooling cycles, which result in the amplification of the target DNA sequence if it is present in the sample [31].

PCR amplification exponentially increases the number of copies of the target DNA sequence, making it easier to detect. The amplified DNA can be visualized by various methods, such as gel electrophoresis or through the use of fluorescent probes. The presence of a specific band or a fluorescent signal indicates the presence of Orientia tsutsugamushi DNA in the sample, confirming the diagnosis of scrub typhus [32].

One of the advantages of PCR testing is its high sensitivity, which allows for the detection of low amounts of bacterial DNA in the clinical sample. This makes PCR particularly useful in the early stages of infection when the bacterial load may be low. Additionally, PCR tests can provide rapid results, making them valuable in clinical settings where prompt diagnosis is



crucial for patient management [33]. However, it's important to note that PCR testing requires specialized laboratory facilities, equipment, and trained personnel. The process involves careful handling of samples and strict adherence to quality control measures to minimize the risk of contamination. The cost of PCR testing may also be higher compared to serological assays like IFA or ELISA.

#### Weil-Felix Test:

The Weil-Felix test is a diagnostic test that was developed several decades ago to detect antibodies against various strains of rickettsial bacteria, including Orientia tsutsugamushi [13], [15]. The test is based on the phenomenon of cross-reactivity between rickettsial antigens and antigens derived from certain strains of Proteus bacteria. The test utilizes a mixture of Proteus antigens to detect antibodies produced by the patient's immune system in response to rickettsial infection [34], [35].

The Weil-Felix test involves collecting a blood sample from the patient and mixing it with the Proteus antigen solution. If the patient has been exposed to Orientia tsutsugamushi or other rickettsial bacteria, antibodies against these pathogens may cross-react with the Proteus antigens [36], [37], leading to agglutination or clumping of the bacterial suspension. Agglutination is observed macroscopically or microscopically, indicating the presence of antibodies against rickettsial bacteria. However, despite its historical significance, the Weil-Felix test has limitations in terms of sensitivity and specificity compared to more modern diagnostic tests. Cross-reactivity with non-rickettsial bacterial infections and other factors can lead to false-positive results, reducing the test's specificity [38]. Additionally, the test may not always detect antibodies in the early stages of infection when the antibody levels are low.

As a result, the Weil-Felix test is less commonly used nowadays, especially in regions where more accurate and specific diagnostic methods, such as IFA, ELISA, or PCR, are available. These newer tests offer higher sensitivity and specificity, allowing for more accurate and reliable detection of Orientia tsutsugamushi infections. They have largely replaced the Weil-Felix test in routine clinical practice.

## **Conclusion**

Scrub typhus is an infectious disease caused by Orientia tsutsugamushi and transmitted through mite bites. Its wide distribution and potential complications make it a significant public health concern in endemic regions.

The Indirect Immunofluorescence Assay (IFA) is a valuable diagnostic tool for detecting antibodies against Orientia tsutsugamushi in patients suspected of having scrub typhus. Its high sensitivity and ability to assess changes in antibody titer between acute and convalescent samples make it the gold standard for scrub typhus diagnosis. Despite its technical requirements and the possibility of false results, the IFA test remains an essential and reliable method in clinical laboratories for confirming the presence of antibodies against Orientia tsutsugamushi and aiding in the timely management of scrub typhus cases.

ELISA is a widely used serological test for the detection of antibodies against Orientia tsutsugamushi. It offers a cost-effective and scalable alternative to IFA, although it may have slightly lower sensitivity and specificity. ELISA is particularly useful for large-scale screening and can detect antibodies at different stages of infection. Despite its limitations, ELISA



remains a valuable tool in the diagnosis and surveillance of scrub typhus, contributing to timely and efficient management of the disease.

PCR tests are highly sensitive and specific methods for directly detecting Orientia tsutsugamushi DNA in clinical samples. They are particularly useful in the early stages of infection when antibody levels may be low. PCR testing requires specialized laboratory facilities and equipment, making it more suitable for centralized diagnostic laboratories. Despite these requirements, PCR remains an invaluable tool in the diagnosis and surveillance of scrub typhus, enabling timely detection and appropriate management of patients.

The Weil-Felix test is an older diagnostic test for detecting antibodies against rickettsial bacteria, including Orientia tsutsugamushi. However, it has limitations in terms of sensitivity and specificity compared to more modern tests. Due to the availability of more accurate diagnostic methods, the Weil-Felix test is now less commonly used in clinical settings. Healthcare professionals rely on more advanced techniques, such as IFA, ELISA, or PCR, for the accurate diagnosis of scrub typhus caused by Orientia tsutsugamushi.

### References

- [1] C.-C. Wang, S.-F. Liu, J.-W. Liu, Y.-H. Chung, M.-C. Su, and M.-C. Lin, "Acute respiratory distress syndrome in scrub typhus," *Am. J. Trop. Med. Hyg.*, vol. 76, no. 6, pp. 1148–1152, Jun. 2007.
- [2] S. Palanivel, K. Nedunchelian, and V. Poovazhagi, "Clinical profile of scrub typhus in children," *The Indian Journal of*, 2012.
- [3] K. Silpapojakul, "Scrub typhus in the Western Pacific region," *Ann. Acad. Med. Singapore*, 1997.
- [4] S. K. Mahajan, J. M. Rolain, and R. Kashyap, "Scrub typhus in Himalayas," *Emerg. Infect. Dis.*, 2006.
- [5] N. Patil *et al.*, "Spot the dot: solve the mystery: tsutsugamushi disease," *Res. J. Pharm. Biol. Chem. Sci.*, vol. 7, no. 1, pp. 1752–1755, 2016.
- [6] T. Weitzel, S. Dittrich, J. López, and W. Phuklia, "Endemic scrub typhus in South America," *England Journal of ...*, 2016.
- [7] S. Rajapakse and P. Weeratunga, "Clinical manifestations of scrub typhus," *Transactions* of the, 2017.
- [8] S. Chakraborty and N. Sarma, "Scrub typhus: an emerging threat," *Indian J. Dermatol.*, 2017.
- [9] D. Kim, K. J. Won, C. Y. Park, K. D. Yu, and H. S. Kim, "Distribution of eschars on the body of scrub typhus patients: a prospective study," *The American journal*, 2007.
- [10] D.-M. Kim, S. W. Kim, S.-H. Choi, and N. R. Yun, "Clinical and laboratory findings associated with severe scrub typhus," *BMC Infect. Dis.*, vol. 10, no. 1, pp. 1–7, Apr. 2010.
- [11] M. Vivekanandan, A. Mani, Y. S. Priya, A. P. Singh, S. Jayakumar, and S. Purty, "Outbreak of scrub typhus in Pondicherry," *J. Assoc. Physicians India*, vol. 58, pp. 24–28, Jan. 2010.
- [12] V. Anitharaj *et al.*, "Serological Diagnosis of Acute Scrub Typhus in Southern India: Evaluation of InBios Scrub Typhus Detect IgM Rapid Test and Comparison with other Serological Tests," *J. Clin. Diagn. Res.*, vol. 10, no. 11, pp. DC07-DC10, Nov. 2016.
- [13] M. Koraluru, I. Bairy, M. Varma, and S. Vidyasagar, "Diagnostic validation of selected serological tests for detecting scrub typhus," *Microbiol. Immunol.*, vol. 59, no. 7, pp. 371– 374, Jul. 2015.
- [14] S. K. Mahajan, "Scrub typhus," JAPI, vol. 53, no. 955, p. 269, 2005.



- [15] J. A. J. Prakash, O. C. Abraham, and E. Mathai, "Evaluation of tests for serological diagnosis of scrub typhus," *Trop. Doct.*, vol. 36, no. 4, pp. 212–213, Oct. 2006.
- [16] S. Stephen, D. Gunasekaran, and V. Anitharaj, "Application of Enzyme Linked Immunosorbent Assay (ELISA) and Indirect Fluorescent Antibody Test for Serodiagnosis of Acute Scrub Typhus in and Around Puducherry, India," J. Clin. Diagn. Res., vol. 12, no. 11, 2018.
- [17] M. L. Clements, J. S. Dumler, P. Fiset, C. L. Wisseman Jr, M. J. Snyder, and M. M. Levine, "Serodiagnosis of Rocky Mountain spotted fever: comparison of IgM and IgG enzymelinked immunosorbent assays and indirect fluorescent antibody test," J. Infect. Dis., vol. 148, no. 5, pp. 876–880, Nov. 1983.
- [18] S. D. Blacksell *et al.*, "Optimal cutoff and accuracy of an IgM enzyme-linked immunosorbent assay for diagnosis of acute scrub typhus in northern Thailand: An alternative reference method to the IgM immunofluorescence assay," *J. Clin. Microbiol.*, vol. 54, no. 6, pp. 1472–1478, Jun. 2016.
- [19] S. Zhang *et al.*, "Scrub typhus in previously unrecognized areas of endemicity in China," *J. Clin. Microbiol.*, vol. 48, no. 4, pp. 1241–1244, Apr. 2010.
- [20] D.-M. Kim *et al.*, "Usefulness of nested PCR for the diagnosis of scrub typhus in clinical practice: A prospective study," *Am. J. Trop. Med. Hyg.*, vol. 75, no. 3, pp. 542–545, Sep. 2006.
- [21] R. Phetsouvanh, S. D. Blacksell, K. Jenjaroen, N. P. J. Day, and P. N. Newton, "Comparison of indirect immunofluorescence assays for diagnosis of scrub typhus and murine typhus using venous blood and finger prick filter paper blood spots," *Am. J. Trop. Med. Hyg.*, vol. 80, no. 5, pp. 837–840, May 2009.
- [22] J. Pradutkanchana, K. Silpapojakul, H. Paxton, S. Pradutkanchana, D. J. Kelly, and D. Strickman, "Comparative evaluation of four serodiagnostic tests for scrub typhus in Thailand," *Trans. R. Soc. Trop. Med. Hyg.*, vol. 91, no. 4, pp. 425–428, Jul-Aug 1997.
- [23] C. S. Eisemann and J. V. Osterman, "Antigens of scrub typhus rickettsiae: separation by polyacrylamide gel electrophoresis and identification by enzyme-linked immunosorbent assay," *Infect. Immun.*, vol. 32, no. 2, pp. 525–533, May 1981.
- [24] J. Jiang *et al.*, "Laboratory diagnosis of two scrub typhus outbreaks at Camp Fuji, Japan in 2000 and 2001 by enzyme-linked immunosorbent assay, rapid flow assay, and Western blot assay using outer membrane 56-kD recombinant proteins," *Am. J. Trop. Med. Hyg.*, vol. 69, no. 1, pp. 60–66, 2003.
- [25] P. N. D. Elders *et al.*, "Diagnostic accuracy of an in-house Scrub Typhus enzyme linked immunoassay for the detection of IgM and IgG antibodies in Laos," *PLoS Negl. Trop. Dis.*, vol. 14, no. 12, p. e0008858, Dec. 2020.
- [26] R. Valsalan, K. Kosaraju, T. Sohanlal, and P. S. P. Kumar, "Hemophagocytosis in scrub typhus," *J. Postgrad. Med.*, vol. 56, no. 4, pp. 301–302, Oct-Dec 2010.
- [27] D. J. Kelly, D. P. Marana, C. K. Stover, E. V. Oaks, and M. Carl, "Detection of Rickettsia tsutsugamushi'by Gene Amplification Using Polymerase Chain Reaction Techniques," NAVAL MEDICAL RESEARCH INST BETHESDA MD, 1990.
- [28] S. T. Tay, S. Nazma, and M. Y. Rohani, "Diagnosis of scrub typhus in Malaysian aborigines using nested polymerase chain reaction," *Southeast Asian J. Trop. Med. Public Health*, vol. 27, no. 3, pp. 580–583, Sep. 1996.
- [29] Y. Sugita, Y. Yamakawa, K. Takahashi, T. Nagatani, K. Okuda, and H. Nakajima, "A polymerase chain reaction system for rapid diagnosis of scrub typhus within six hours," *Am. J. Trop. Med. Hyg.*, vol. 49, no. 5, pp. 636–640, Nov. 1993.



- [30] Y. Sugita, T. Nagatani, K. Okuda, Y. Yoshida, and H. Nakajima, "Diagnosis of typhus infection with Rickettsia tsutsugamushi by polymerase chain reaction," J. Med. Microbiol., vol. 37, no. 5, pp. 357–360, Nov. 1992.
- [31] S. Sankar et al., "Detection of scrub typhus by real-time polymerase chain reaction and immunoglobulin M ELISA among patients with acute febrile illness," J. Nat. Sci. Biol. Med., vol. 11, no. 1, p. 66, 2020.
- [32] Y. Furuya, Y. Yoshida, T. Katayama, S. Yamamoto, and A. Kawamura Jr, "Serotype-specific amplification of Rickettsia tsutsugamushi DNA by nested polymerase chain reaction," J. *Clin. Microbiol.*, vol. 31, no. 6, pp. 1637–1640, Jun. 1993.
- [33] F. Kawamori *et al.*, "Two-step polymerase chain reaction for diagnosis of scrub typhus and identification of antigenic variants of Rickettsia tsutsugamushi," *J. Vet. Med. Sci.*, vol. 55, no. 5, pp. 749–755, Oct. 1993.
- [34] G. W. Brown, D. M. Robinson, D. L. Huxsoll, T. S. Ng, and K. J. Lim, "Scrub typhus: a common cause of illness in indigenous populations," *Trans. R. Soc. Trop. Med. Hyg.*, vol. 70, no. 5–6, pp. 444–448, 1976.
- [35] A. Felix, "Technique and interpretation of the Weil-Felix test in typhus fever," *Trans. R. Soc. Trop. Med. Hyg.*, vol. 37, no. 5, pp. 321–341, Mar. 1944.
- [36] J. Manosroi, S. Chutipongvivate, W. Auwanit, and A. Manosroi, "Early diagnosis of scrub typhus in Thailand from clinical specimens by nested polymerase chain reaction," *Southeast Asian J. Trop. Med. Public Health*, vol. 34, no. 4, pp. 831–838, Dec. 2003.
- [37] S. K. Mahajan, R. Kashyap, A. Kanga, V. Sharma, B. S. Prasher, and L. S. Pal, "Relevance of Weil-Felix test in diagnosis of scrub typhus in India," *J. Assoc. Physicians India*, vol. 54, pp. 619–621, Aug. 2006.
- [38] K. Amano, N. Suzuki, M. Fujita, Y. Nakamura, and T. Suto, "Serological reactivity of sera from scrub typhus patients against Weil-Felix test antigens," *Microbiol. Immunol.*, vol. 37, no. 12, pp. 927–933, 1993.